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FABRICATION OF LARGE AREA POLYMER MICROFILTERS VIA VACUUM ASSISTED UV MICRO-MOLDING



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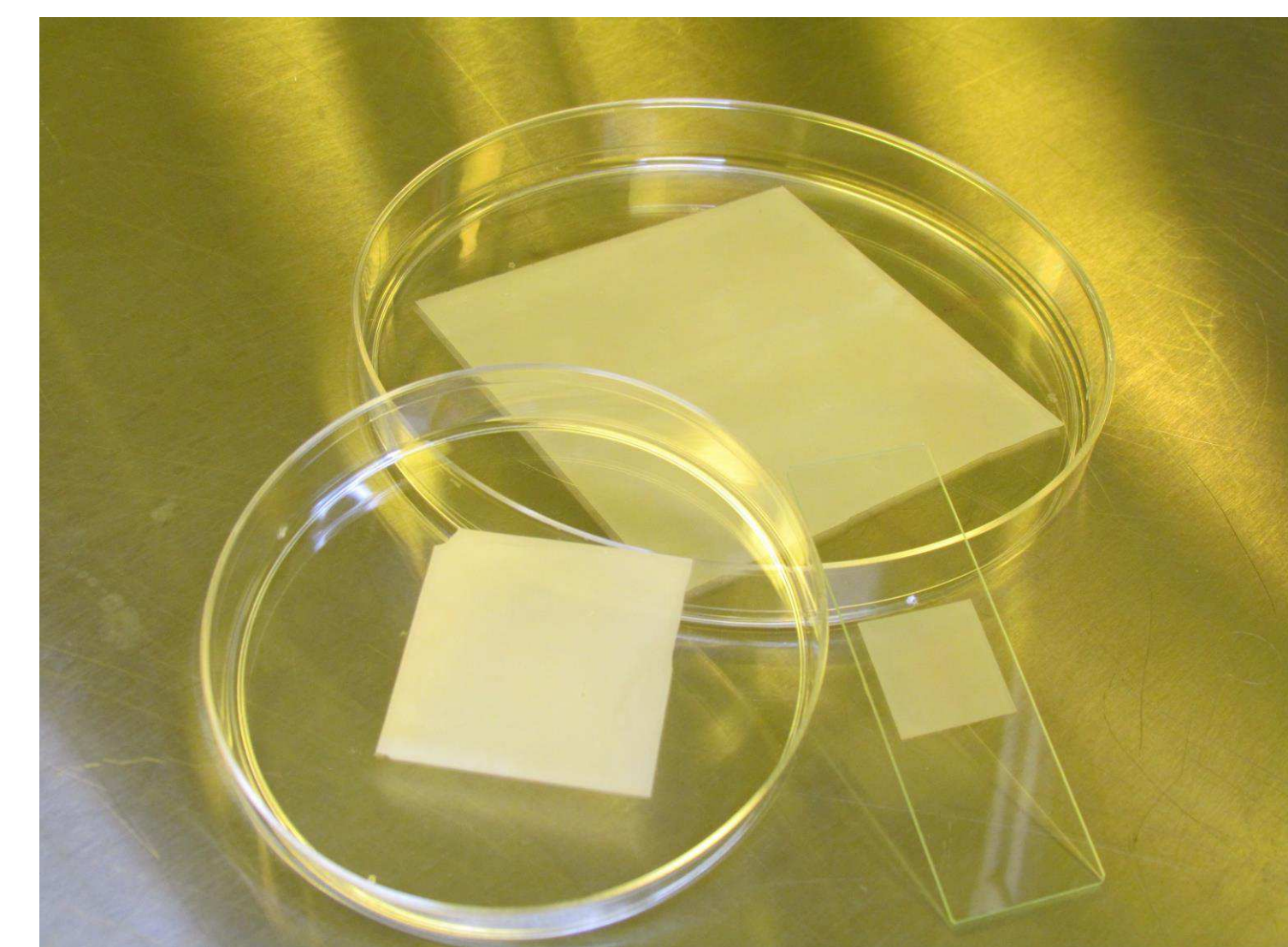
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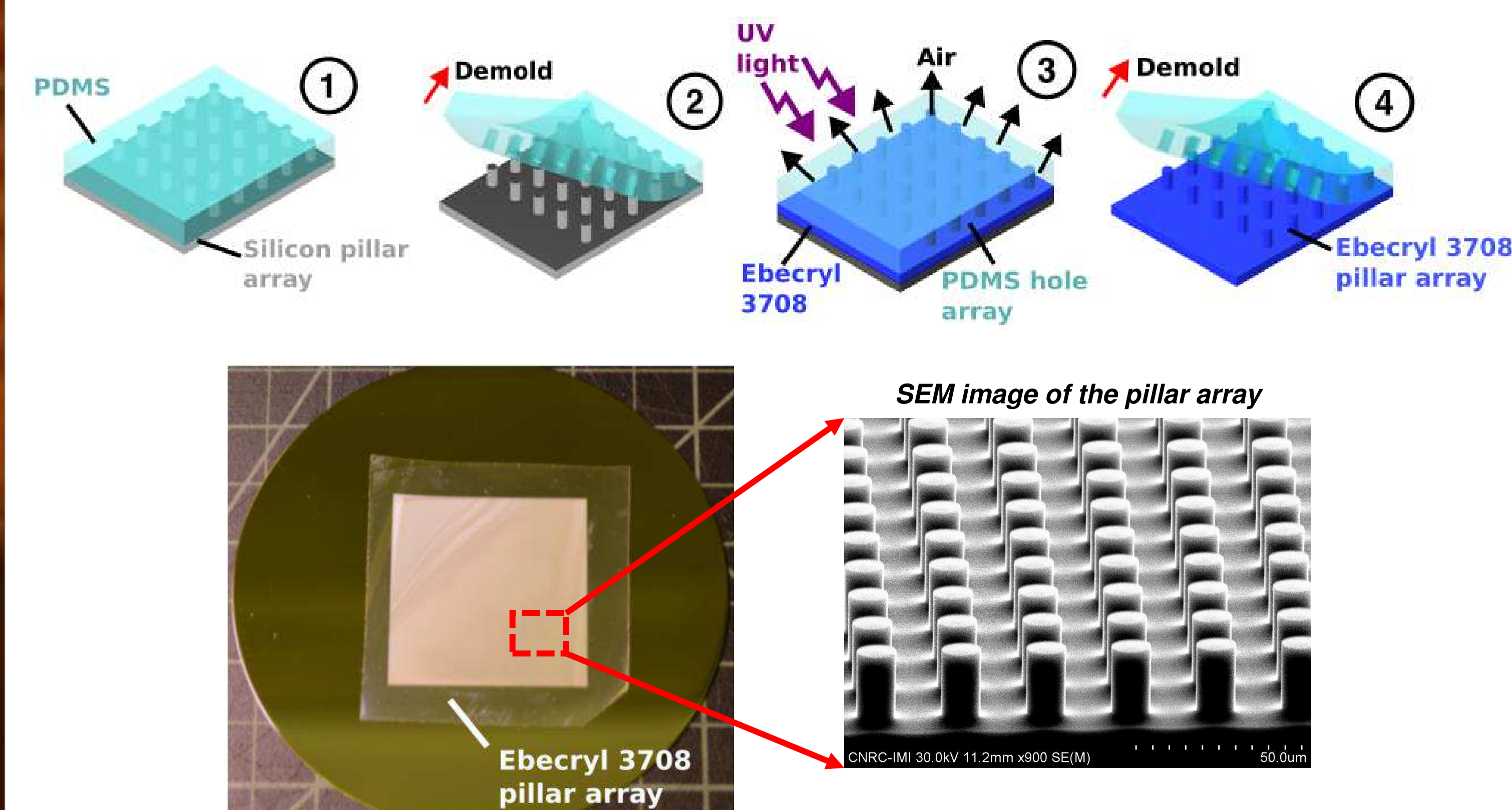
Introduction

- ❖ Microfilters (MFs) with pore openings from $\sim 1 - 100 \mu\text{m}$ can be used for microbead isolation, CTC capture, cell population enrichment, sample preparation and purification, among others.
- ❖ Many different MFs exist, but they suffer from drawbacks, notably:
 - Silicon^[1] and parylene C^[2] MFs can be made precisely, but fabrication is cumbersome. Also, Si is opaque and brittle, while parylene C is auto-fluorescent.
 - Polymer track-etched membranes^[3] are available commercially, but porosity is limited to values $< 30\%$, which limits filtration flow rate.
- ❖ Here, we introduce (i) a robust, low cost, and scalable fabrication process based on vacuum assisted micro-molding^[4] and UV-curable polymers for making (ii) large area, high porosity, transparent MFs with pore sizes ranging from $3 - 50 \mu\text{m}$.



Microfilter mold fabrication

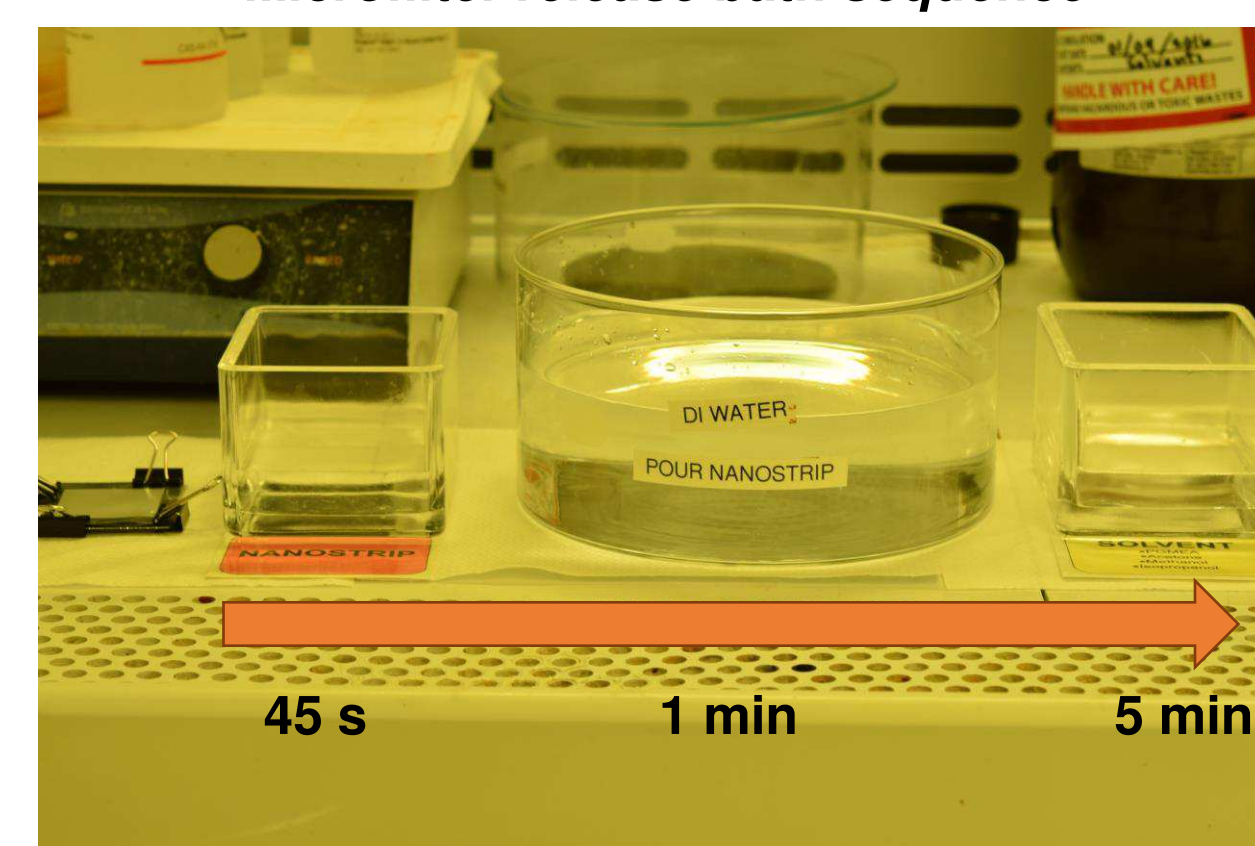
- ❖ A silicon master mold is fabricated through standard photolithography techniques.
- ❖ The mold is replicated twice, first in PDMS, and then in a UV-curable epoxy (Ebecryl® 3708).



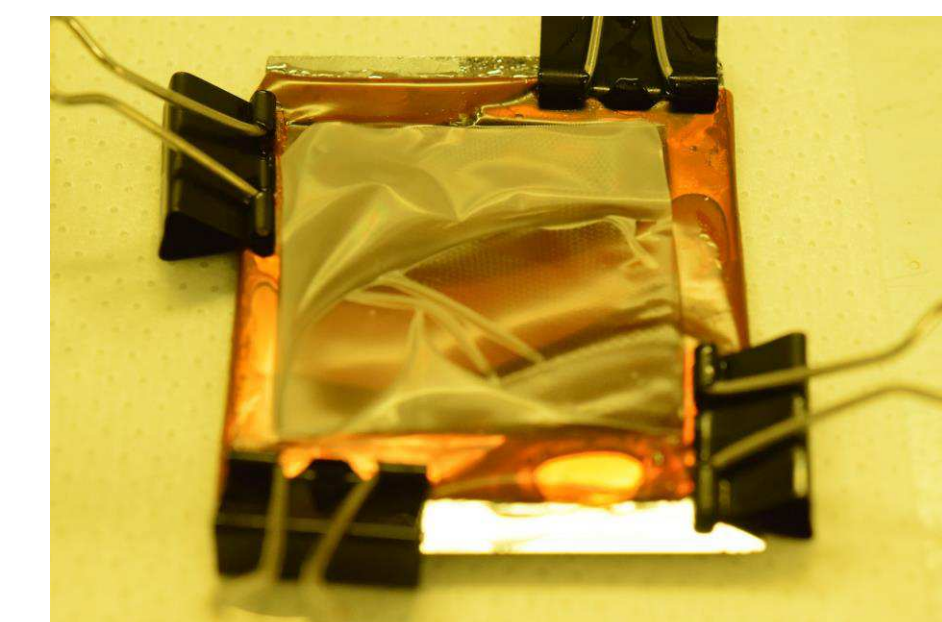
Microfilter release

- ❖ Once the pre-polymer has filled the entire mold, it is exposed to 3 min of UV radiation. The cover layer is then peeled off.
- ❖ The mold is subjected to a 45 s acid bath (Nano-Strip 2X®), followed by a 1 min DI water wash, and a 5 min acetone bath, which leads to spontaneous release of the free-standing microfilter membrane.

Microfilter release bath sequence



Microfilter detached from mold

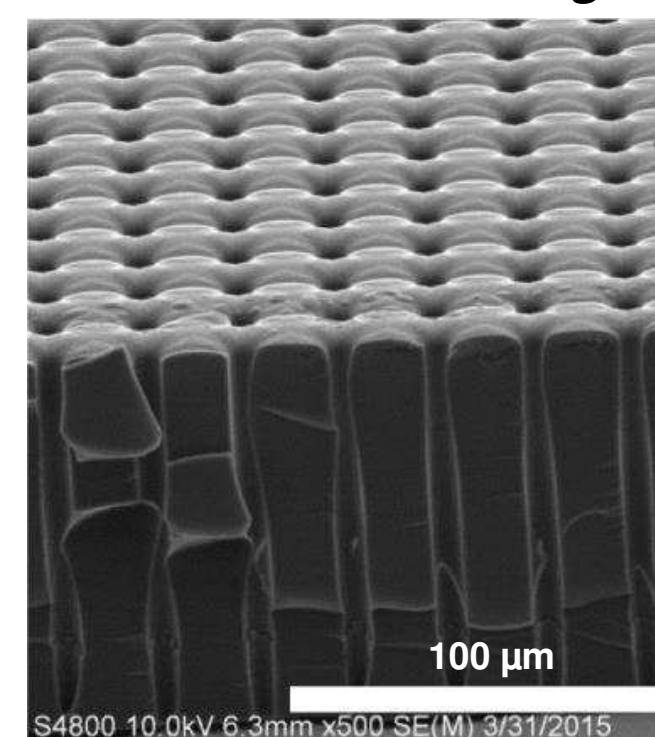


After the filters demold they can be easily picked with tweezers. They are cleaned with methanol and dried under a nitrogen stream.

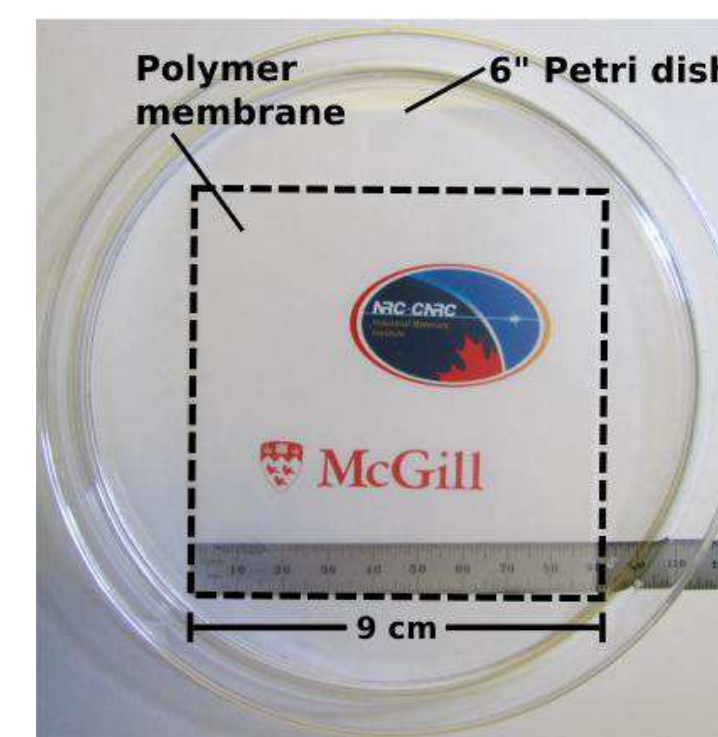
Microfilters

- ❖ Microfilters with thickness from 8 to $100 \mu\text{m}$, size from 4×4 to $9 \times 9 \text{ cm}^2$, pore size from 3 to $50 \mu\text{m}$, different pore shapes, and porosity as high as $\sim 60\%$ have been successfully fabricated using the proposed method.

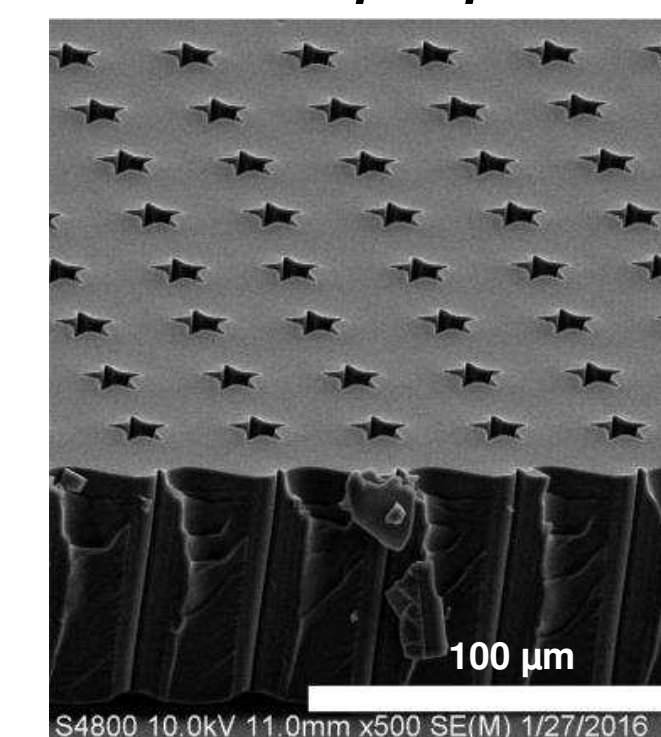
Microfilter SEM image



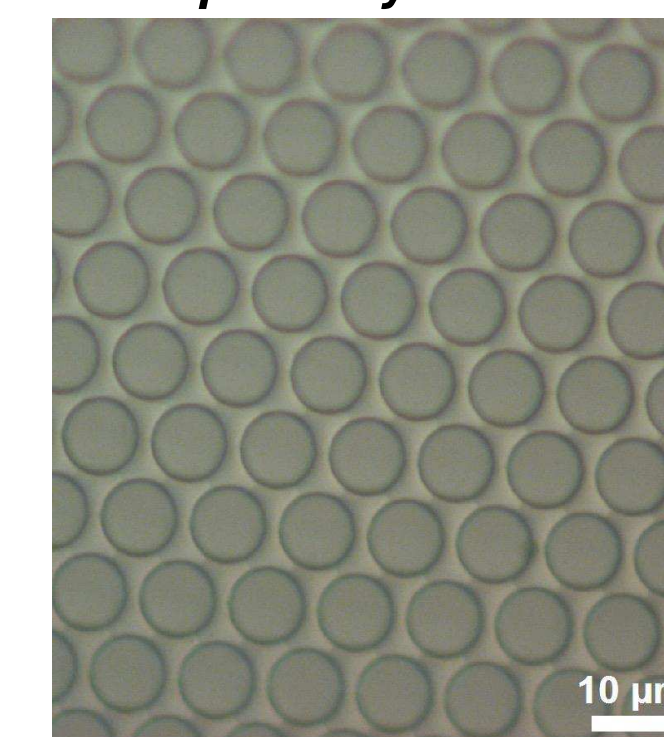
9x9 cm² microfilter



Star shaped pores

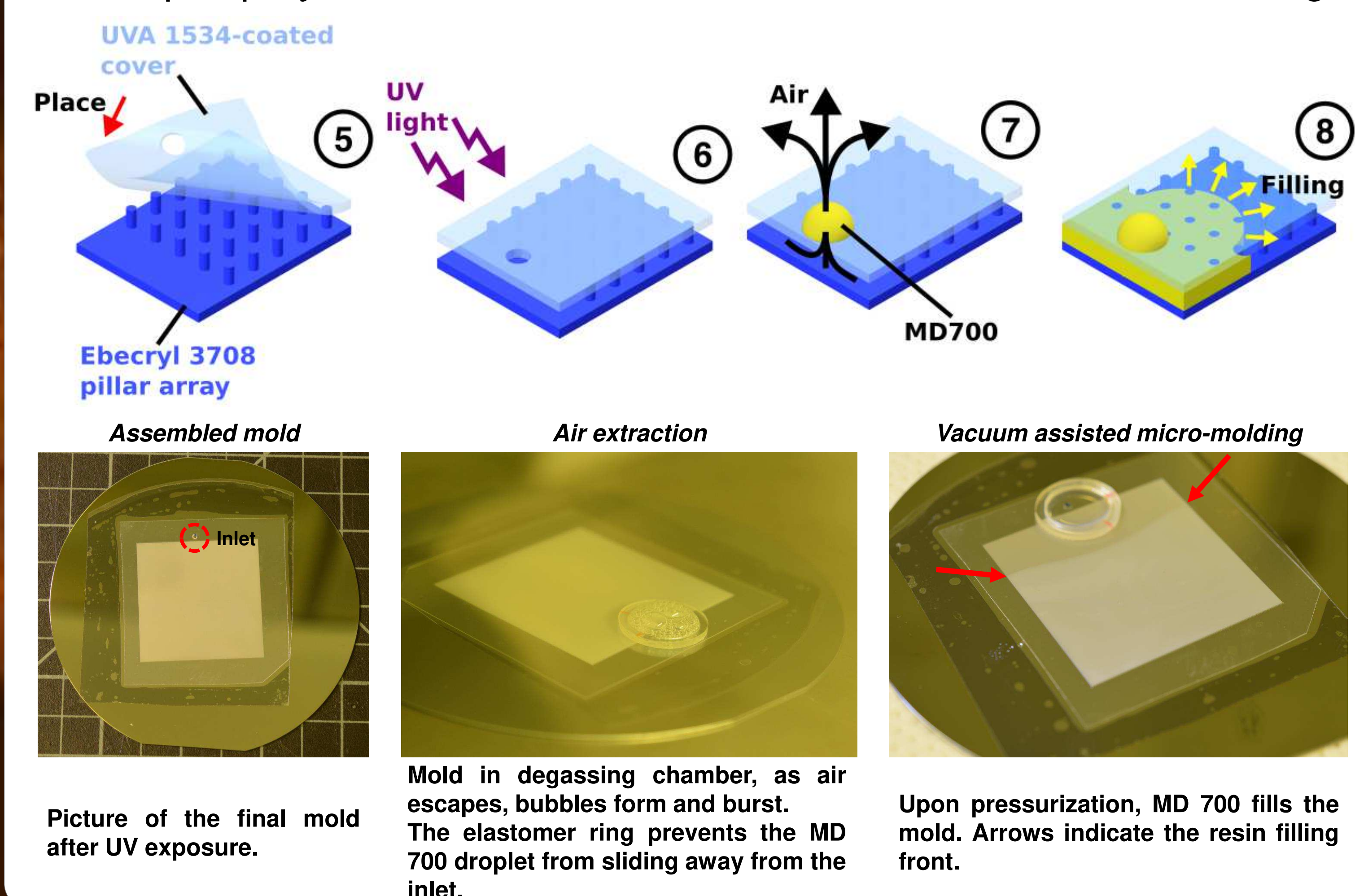


60% porosity microfilter



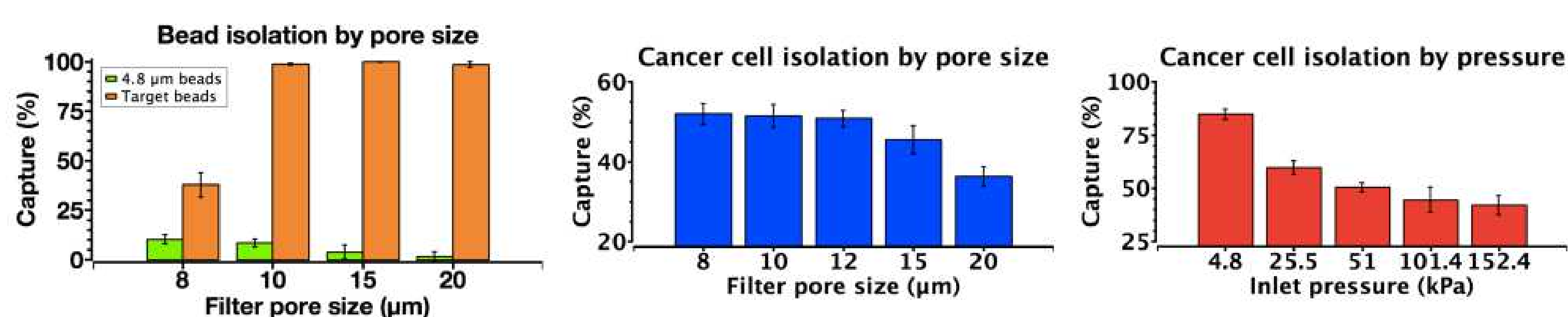
Fluoropolymer filling

- ❖ The Ebecryl® 3708 mold is covered using a PET film coated with a different UV-curable resin (UVA 1534). A single opening will be used to fill in the pre-polymer.
- ❖ A droplet of a fluorinated methacrylate pre-polymer (Fluorolink MD 700), with a dynamic viscosity of 430 cP , is placed on the inlet.
- ❖ The pre-polymer fills the mold via vacuum assisted micro-molding.



Microfiltration experiments

- ❖ The MFs were used to filter microbeads ($8 - 20 \mu\text{m}$) and MDA-MB-231 cancer cells suspended in PBS. Filtration performance was analyzed by characterizing pre- and post-filtration samples using flow cytometry.
- ❖ The MFs are essentially defect-free, rigid ($E=10 \text{ MPa}$), and tough, enabling filtration at high flow rates and without failure.
- ❖ Separation of CTC-like cells from blood is reported elsewhere.^[5]



Conclusions

- ❖ We present a powerful method for **fabrication of robust and transparent polymer MFs** with pores down to $3 \mu\text{m}$, porosity up to 60% , and filter size up to $9 \times 9 \text{ cm}^2$ large.
- ❖ The MFs were used for bead and cancer cell isolation.
- ❖ **The MFs are cheap, tough, have a precise pore size, are easy to manipulate and cut to size, chemically resistant and compatible with chemical functionalization**, which collectively should make them useful for a wide range of chemical and biological applications.

Acknowledgements



References

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