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Hounsfield unit recovery in clinical cone beam CT images of the thorax acquired for image guided radiation therapy

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Abstract

A comprehensive artefact correction method for clinical cone beam CT (CBCT) images acquired for image guided radiation therapy (IGRT) on a commercial system is presented. The method is demonstrated to reduce artefacts and recover CT-like Hounsfield units (HU) in reconstructed CBCT images of five lung cancer patients.

Projection image based artefact corrections of image lag, detector scatter, body scatter and beam hardening are described and applied to CBCT images of five lung cancer patients. Image quality is evaluated through visual appearance of the reconstructed images, HU-correspondence with the planning CT images, and total volume HU error.

Artefacts are reduced and CT-like HUs are recovered in the artefact corrected CBCT images. Visual inspection confirms that artefacts are indeed suppressed by the proposed method, and the HU root mean square difference between reconstructed CBCTs and the reference CT images are reduced by 31% when using the artefact corrections compared to the standard clinical CBCT reconstruction.

A versatile artefact correction method for clinical CBCT images acquired for IGRT has been developed. HU values are recovered in the corrected CBCT



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images. The proposed method relies on post processing of clinical projection images, and does not require patient specific optimisation. It is thus a powerful tool for image quality improvement of large numbers of CBCT images.

Keywords: cone beam CT, image guided radiotherapy, image quality, Monte Carlo

(Some figures may appear in colour only in the online journal)

1. Introduction

As cone beam CT (CBCT) imaging for image guided radiation therapy (IGRT) is becoming standard practice in many centres, the potential use of CBCT imaging as a basis for personalised radiotherapy is proposed in numerous ways. Dose calculation and accumulation has been proposed by many studies, with online adaptive replanning as the most challenging and desirable application (Yang *et al* 2007, Rong *et al* 2010, Fotina *et al* 2012, Elstrøm *et al* 2014, Vestergaard *et al* 2014).

For lung cancer patients in particular, anatomical changes occur frequently, requiring treatment plan adaptation to deliver the intended radiation dose to the target while sparing the organs at risk (Møller *et al* 2014). Such adaptations are labour intensive, and it is desirable to be able to use the CBCT images acquired for IGRT to either perform the plan adaptation and replanning, or determine for which patients a new CT scan and replanning procedure is required dosimetrically. Another more recent application of CBCT images of the thoracic region is the extraction of biomarkers to predict tumour control or toxicity measures during treatment of lung cancer, with a view to differentiating the patient population based on their radiosensitivity (Bertelsen *et al* 2011, Brink *et al* 2014, Jabbour *et al* 2015, Bernchou *et al* 2015). Such differentiation has the potential of providing better tumour control rates for those patients who can tolerate higher dose levels, while providing a better quality of life for those patients who are prone to severe toxic sideeffects such as pneumonitis and pulmonary fibrosis.

The main limiting factor for all of the above mentioned extended uses of CBCT images is the image quality. CBCT image quality has been studied since the first gantry mounted CBCT scanner was realised (Jaffray *et al* 2002), with five different factors known as the main causes for the limited image quality, namely image lag (Siewerdsen and Jaffray 1999), detector scatter (Poludniowski *et al* 2011), body scatter (Siewerdsen and Jaffray 2001), beam hardening (Herman 1979), and a potential truncated field of view (Ohnesorge *et al* 2000). Each factor has been studied extensively using different phantoms, but to our knowledge no study has yet investigated the potential increased image quality when all artefact sources are considered together in relation to clinical CBCT imaging of patients. This study investigates how clinical CBCT images of the chest can be improved by correcting for all five artefact sources. Post-processing of raw image data from the Elekta XVI CBCT system is performed, and we demonstrate that proper Hounsfield Units (HUs) can be recovered from the existing projection images.

2. Materials and methods

All CBCT scans were acquired on the Elekta XVI R4.5 system, either mounted on an Elekta Versa HD or an Elekta Synergy accelerator (Elekta Ltd, Crawley, UK). The XVI R4.5 CBCT system is based on a Dunlee type DA1094/DU694 x-ray tube (Dunlee, IL, USA) and a PerkinElmer XRD 1640 flat panel imager (PerkinElmer Inc, MA, USA).



Figure 1. Overview of the image processing and reconstruction for each of the four different CBCT reconstructions. The *XVI* method is the clinical reference method. *RTK(XVI)* resembles the *XVI* method, but uses RTK to perform the image reconstruction. *RTK(BS)* uses Monte Carlo based scatter correction and RTK reconstruction, while *RTK(All)* includes correction for image lag, detector scatter, MC based body scatter correction, and beam hardening correction prior to reconstruction with RTK.

Five lung cancer patients were chosen from our clinical database, selected to have had a CBCT scan and a re-simulation CT (rCT) scan performed on the same day to facilitate a comparison between CBCT and CT image quality. CBCT scans were acquired using the small field of view without bowtie filtration, allowing a cylindrical volume of length 276.6 mm and diameter 270 mm to be reconstructed. For each scan, approximately 750 frames were acquired over an arc of 220°, using our clinical protocol for fast 4D CBCT imaging. Each frame was exposed to 0.32 mAs, and acquired at a fixed frame rate of 5.5 Hz. The CBCT images were acquired as 4D scans in the clinic, but were reconstructed in 3D in the present work. All patients were scanned isocentrically, with only small (≤ 1 cm) couch shifts applied prior to treatment. In the following sections, all the applied image corrections will be described briefly to provide the full overview of artefact corrections.

2.1. CBCT reconstructions

A total of four reconstructions were made from each CBCT scan. A schematic drawing of the image processing associated with each of the four reconstructions is shown in figure 1.

2.1.1. XVI. For reference, a CBCT reconstruction was made using our standard clinical preset for 3D reconstruction in the XVI software. This uses a simple scatter correction, where the scatter in each projection image is estimated as a fraction of the mean signal in all pixels with a reading of less than 32×10^3 . In our clinic, the scatter subtraction was found to be optimal at 20% of the mean signal in each projection. If the scatter correction results in pixel values less than 20, a constant is added to the entire projection image to ensure the minimum value is 20. The XVI projection images are stored as 16 bit integer images, thus containing 2^{16} gray levels. Following the scatter correction, XVI uses a standard FDK-type filtered backprojection algorithm (Feldkamp *et al* 1984) to reconstruct the CBCT images. No information about the use of truncation correction in the XVI system was available. This clinical reconstruction will be denoted XVI.

2.1.2. RTK(XVI). To investigate the effects of changing the reconstruction algorithm from the XVI native to the FDK-type algorithm provided with the reconstruction toolkit (RTK) (Rit *et al* 2014), a reconstruction was made using a similar scatter correction method as that in the XVI software prior to reconstruction. Furthermore, a 2D median filter with a

 5×5 pixel kernel was applied to the projection images prior to reconstruction, similar to what is used inherently by XVI. The use of RTK allows for truncation correction based on the work of Ohnesorge *et al* (2000), which was used to approximate the truncated anatomy as going to zero over a distance of 20% of the projection image size. Additionally, a Hann window set to a cut-off frequency of 0.8 was used to reduce high frequency noise in the reconstructed images. This reconstruction is referred to as *RTK(XVI)*, and was included in the comparison to emphasise the difference in image quality resulting from changing the reconstruction algorithm only, while keeping image corrections similar to the clinical implementation in XVI.

2.1.3. RTK(BS). With the main reason for poor CBCT image quality being body scatter from the patient, a reconstruction was made using an MC based scatter correction method developed from our previously published model (Thing et al 2013). The EGSnrc (Kawrakow 2000, Kawrakow et al 2011) user code egs cbct (Mainegra-Hing and Kawrakow 2008, 2010) was used to provide fast MC simulations of patient specific scatter distributions, taking advantage of the variance reduction techniques (VRTs) implemented in egs cbct (Thing and Mainegra-Hing 2014). To provide accurate scatter estimates, the full x-ray source was simulated using a compiled BEAM source in egs cbct (Rogers et al 2011). Specifications of the x-ray tube and collimation were found in manuals and data sheets of the Dunlee tube and the XVI system, and details of the source simulation is provided in appendix A. Photon interaction cross sections from the XCOM compilation (Hubbell and Seltzer 2004) were used, with Rayleigh scattering included in the simulations. Photon low energy cut-off was set to 1 keV in the entire simulation geometry, and to avoid spending time simulating electrons which do not reach the detector, the electron low energy cut off was set to 1 MeV after the primary collimation and filtration of x-rays from the BEAM source.

In addition to the scatter calculations, primary photons were simulated to allow an appropriate normalisation between simulated and clinical projection images. The primary simulation was performed using a spatially uniform source with an energy distribution obtained from the full MC simulation of the XVI source. To facilitate fast calculations, a ray tracing mode was implemented in egs_cbct and used for the primary calculations. All simulated projection images were normalised to open field simulations.

The energy response of the XVI flat panel detector was adopted from Roberts *et al* (2008). In the present work, the full detector resolution of 512×512 quadratic pixels with a side length of 0.8 mm was used, and projection images were simulated at an angular interval of 2.5°.

Patient specific primary and scatter simulations were allowed through the creation of a simulation phantom based on the planning CT image for each patient. Four different tissues were assigned based on the HU in each voxel of the planning CT, namely air, lung tissue, soft tissue, and bone. Within each tissue, the density varies according to the exact HU found in each CT voxel. With the simulations based on the planning CT, it is possible to perform all MC calculations prior to CBCT acquisition.

Based on the assumption that the simulated total signal equals the measured clinical projection images when normalised to the appropriate open field signal $(I^{\text{Sim}}/I_0^{\text{Sim}} = I^{\text{Clin}}/I_0^{\text{Clin}})$, the scatter subtraction is obtained through a simple scaling of the simulated scatter signal:

$$I_{\rm MC}^{\rm Clin} = I^{\rm Clin} - K \cdot \frac{I_{\rm scatter}^{\rm Sim}}{I_0^{\rm Sim}}.$$
 (1)

Here, I^{Clin} denotes the measured clinical projection images, and $I_{\text{scatter}}^{\text{Sim}}$ the simulated scatter signal. *K* is equivalent to the average clinical open field signal $\langle I_0^{\text{Clin}} \rangle$; a quantity which is not measured or stored in the XVI system. Therefore, it was estimated retrospectively in the following way: For each set of projection images, a smoothed histogram was calculated for both the clinical and simulated projections. The histograms were created with 100 equidistant bins spanning the full range of numbers in the projection image sets. In these two histograms, two distinct peaks were visible, corresponding to signal attenuated by soft tissue or air. These two peaks were used to perform a two-point calibration of *K* through

$$K = \frac{H_{\text{air}}^{\text{Clin}} - H_{\text{soft tissue}}^{\text{Clin}}}{H_{\text{air}}^{\text{Sim}} - H_{\text{soft tissue}}^{\text{Sim}}}$$
(2)

where H denotes the mean value in the most frequent histogram bins corresponding to air and soft tissue. When determining the scaling factor K through the proposed histogram analysis, the requirement of accurate patient positioning (potentially obtained by rigid or deformable image registration) is relaxed due to the low frequency variation found in the scatter signal (Thing *et al* 2013). Therefore, the isocentric patient setup prior to CBCT acquisition was sufficiently accurate for the MC based body scatter correction to be applied.

While equation (1) is conceptually very simple, it has the potential of leading to negative values of I_{MC}^{Clin} behind highly attenuating anatomy or in the case of severe image noise. To avoid these undesired negative numbers, a smooth cut-off function adapted from Xu *et al* (2015) was used in the present study. A γ parameter is defined through the scatter to total ratio:

$$\gamma = \frac{I_{\text{scatter}}}{I_{\text{total}}} \approx \frac{K \cdot I_{\text{scatter}}^{\text{Sim}} / I_0^{\text{Sim}}}{I^{\text{Clin}}}.$$
(3)

Here, the approximation is used to emphasize measurement noise and non-ideal behaviour in the imaging chain, as well as approximations made in the MC simulation. Based on this definition of γ and a cut-off ratio of $\beta = 0.7$ (determined empirically), the scatter corrected signal $I_{\rm MC}^{\rm Clin}$ was calculated as:

$$I_{\rm MC}^{\rm Clin} = \begin{cases} I^{\rm Clin} \cdot (1-\gamma) & \text{for } 0 \leq \gamma \leq \beta \\ I^{\rm Clin} \cdot \left[(1-\beta) \cdot \exp\left(-\frac{\gamma-\beta}{1-\beta}\right) \right] & \text{for } \gamma > \beta. \end{cases}$$
(4)

This formalism reduces to equation (1) for $\gamma \leq \beta$ (full scatter correction), while for larger scatter to total ratios, the scatter subtraction is gradually decreased such that the remaining signal $I_{\rm MC}^{\rm Clin}$ tends towards zero. Equation (4) is derived from the constraints that $I_{\rm MC}^{\rm Clin}$ and its first derivative in γ must be continuous, and that $I_{\rm MC}^{\rm Clin} \rightarrow 0$ as $\gamma \rightarrow \infty$.

Before reconstruction, the scatter corrected projection images were rescaled by the open field signal estimated through (2):

$$I_{\text{rescaled}}^{\text{Clin}} = \frac{I_{\text{MC}}^{\text{Clin}}}{K} \cdot 2^{16}.$$
(5)

This rescaling is applied since RTK expects the open field signal in projection images to have a digital value of 2^{16} , and a completely blocked signal to have a digital value of 0. The rescaled images were median filtered and reconstructed in RTK as described in the previous *RTK(XVI)* method. This method of body scatter correction and RTK reconstruction is referred to as *RTK(BS)*.

2.1.4. RTK(All). The most sophisticated artefact correction method included five steps prior to reconstruction, each of which is described in the following. The order in which to apply the corrections was determined to first correct for the physical effect which occurs last, and then work backwards from the detector related artefact sources to the patient related artefact sources. Thus, the corrections in applied order were image lag, detector scatter, body scatter, and beam hardening, before projection scaling was performed prior to CBCT reconstruction with truncation correction applied. This comprehensive correction and reconstruction is denoted *RTK(All)*.

Image lag. A linear time invariant image lag correction method was developed based on the works by Hsieh, Starman and Mail (Hsieh *et al* 2000, Mail *et al* 2008, Starman *et al* 2011, 2012). Such a model assumes that some signal will be trapped in the detector and not be read out in the correct frame. This leads to a situation where each frame from the detector contains signal from the previously exposed frames, as well as signal from the current exposure. Furthermore, some signal from the current exposure will be stored and read out in the following frames.

In the present model, the time function of the measured signal I^{meas} from a very short exposure I^{exp} (a delta function) is described as a sum of exponentials:

$$I^{\text{meas}}(t) = I^{\exp}\sum_{i=1}^{n} \frac{a_i}{t_i} \exp\left(\frac{-t}{t_i}\right).$$
(6)

Here, a_i is the amplitude of each exponential term, t_i the decay time of each term, and n is the number of exponentials required to describe the decaying signal. a_i is bound by the constraint that $\sum a_i = 1$, and it is noted that the first exponential in the sum has a large amplitude and short decay time, corresponding to the fact that most of the signal from a delta pulse is read out in the first frame.

For continuous exposures, a set of signal storage variables q_i is introduced, each of which is updated frame by frame. The lag correction model of the time dependent detector readout corrects for increases in signal from the previous frames, as well as a lack of signal from the present frame which is stored for future frames. Given a set of measured projections { $I^{\text{Clin}}(k)$ }, where k denotes the frame number starting at k = 1, the lag corrected signal { $I^{\text{Clin}}_{\text{LagCorr}}(k)$ } is calculated as

$$I_{\text{LagCorr}}^{\text{Clin}}(k) = \frac{I^{\text{Clin}}(k) - \sum_{i=1}^{n} q_i(k) \cdot (1 - \exp(-\Delta k/t_i))}{1 - \sum_{i=1}^{n} a_i \exp(-\Delta k/t_i)}$$
(7)

where Δk is the time between frames. All parameters but Δk were determined experimentally. In the present study, the time was measured in units of the fixed time between frames on the XVI R4.5 system (181 ms), and hence $\Delta k = 1$. q_i is defined recursively with $q_i(k = 1) = 0$ and

$$q_i(k+1) = [I_{\text{LagCorr}}^{\text{Clm}}(k) \cdot a_i + q_i(k)] \cdot \exp(-\Delta k/t_i).$$
(8)

To determine a_i and t_i , a number of calibration measurements were performed using the falling step response method. To obtain the calibration measurements, the XVI unit was operated in service mode where it is possible to have continuous readout of the detector while the x-ray source can be turned on and off independently. Measurements were acquired with the x-ray source turned on before the frame readout was enabled. After readout of approximately 200 exposed frames, the x-ray source was turned off, and the decaying signal from the detector was measurements were acquired with the detector was measured for approximately 400 frames. Such falling step response measurements

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i	a_i	t_i
1	0.955	0.240
2	0.0257	3.10
3	0.0120	19.9
4	0.00735	118

Table 1. Fitted parameters from (9) and used by (7) and (8) for lag correction.

Note: t_i is measured in units of the time between frames (181 ms on XVI R4.5).

were acquired at six different exposure levels between 0.3% and 80.3% of saturation. At each level, measurements were repeated three or five times with 15 minutes interval to reduce residual lag between measurements. The measured falling step response functions were fitted to the following multiexponential model denoted by y_{fit} , which is obtained by solving (7) and (8) for $I_{\text{LagCorr}}^{\text{Clin}}(k \ge k_0) = 0$:

$$y_{\rm fit}(k) = \begin{cases} I^{\rm exp} \cdot \sum_{i=1}^{n} a_i + c & \text{for } k \le k_0 \\ I^{\rm exp} \cdot \sum_{i=1}^{n} a_i \cdot \exp\left(\frac{-(k-k_0)}{t_i}\right) + c & \text{for } k > k_0 \end{cases}$$
(9)

where I^{exp} is the calibration field exposure, a_i denotes the amplitudes for each exponential term, t_i the decay time (in units of time between frames), k_0 is the first frame with the x-ray source turned off, and *c* is a constant introduced to compensate for the detector offset which causes the signal to decay to a non-zero number at infinite time. The *c* parameter is negligible in comparison with the measured clinical projection images, and is thus omitted in the actual processing of the clinical CBCT projection images.

To perform the image lag correction, the a_i and t_i constants determined from (9) were input to equations (7) and (8). We found n = 4 to be the simplest model to describe our calibration measurements, as found through analysis of residuals which did not contain time structure after inclusion of the 4 exponential terms. The fitted model parameters are shown in table 1. During investigation of the calibration measurements, we found no reason to include an exposure dependence in our lag correction method. Calibration measurements were performed with one XVI unit, and the lag correction method was verified to correct images acquired on all the units in our clinic.

Detector scatter. An implementation of the detector scatter correction published by Poludniowski *et al* (2011) was used in the present model to enhance contrast in the reconstructed CBCT images. Based on an edge-spread measurement where a lead slab was placed 30 cm from the detector, an experimentally determined line spread function was fitted to a sum of a Gaussian, a MacDonald, and a Lorentzian function:

$$\mathrm{LSF}^{\mathrm{fit}}(x) = \frac{\alpha_1}{\sqrt{2\pi\beta_1^2}} e^{-x^2/(2\beta_1^2)} + \frac{\alpha_2}{\pi\beta_2^2} |x| K_{\mathrm{I}}\left(\frac{|x|}{\beta_2}\right) + \frac{\alpha_3}{\pi\beta_3} \frac{1}{1 + x^2/\beta_3^2}.$$
 (10)

Here, α_i , β_i are fitting parameters, while K_1 is a MacDonald (modified Bessel) function and x is the distance in the line spread profile from the lead edge. A normalisation constraint is imposed, forcing $\sum_i \alpha_i = 1$.

Only the contribution from the broad Lorentzian function was used for subsequent deconvolution of detector scatter from measured projection images due to high-frequency instabilities in the more narrow Gaussian and MacDonald terms during deconvolution. The discrete function (named glare spread function by Poludniowski *et al*) used for deconvolution of detector scatter was

$$\text{GSF}(i,j) = (1 - \alpha_3)\delta_{i,0}\delta_{j,0} + \frac{\alpha_3 \Delta x \Delta y}{2\pi\beta_3^2} \frac{1}{(1 + r^2(i,j) \cdot \beta_3^{-2})^{3/2}},$$
(11)

where *i*, *j* are pixel indices, $\delta_{i,0}$ and $\delta_{j,0}$ are Kronecker deltas, $\Delta x \Delta y$ is the area of a pixel, $r(i,j) = \sqrt{x^2(i) + y^2(j)}$ is the distance from the center of the kernel, and α_3, β_3 are fitted parameters from (10). For the full description on how to perform the calibration measurements, the reader is referred to Poludniowski *et al* (2011).

We found that $\alpha_3 = 8.431 \times 10^{-2}$ and $\beta_3 = 9.291$ provided a good fit across our six clinical XVI systems, with the variations between systems being similar to the variation between repeat measurements on one system. The digital value in pixel *i*, *j* ($I_{\text{DetectorCorr}}^{\text{Clin}}(i, j)$) was determined through Fast Fourier Transforms (FFT) in the following deconvolution:

$$I_{\text{DetectorCorr}}^{\text{Clin}}(i,j) = \text{FFT}^{-1} \left(\frac{\text{FFT}(I^{\text{Clin}}(i,j))}{\text{FFT}(\text{GSF}(i,j))} \right).$$
(12)

To avoid edge effects of the Fourier transforms, all images were zero-padded using a Tukey window to create a smooth transition to zero (Tukey 1967).

Body scatter. The body scatter correction described under the *RTK*(*BS*) method was used to correct for scatter in the comprehensive case as well.

Beam hardening. Beam hardening arises from the fact that CBCT x-ray sources create a spectrum of x-ray energies which are incident on the patient. In the patient, different tissues have different energy dependent attenuation coefficients, and hence the attenuation of the x-ray spectrum will depend on the depth and pathway of the x-rays through the patient. To account for beam hardening, the method proposed by Herman (1979) was adopted. Using the same MC simulation setup to estimate scattered radiation, the patient specific attenuation from primary x-rays was calculated using both the real polyenergetic x-ray source ((μx)_{poly}) and a monoenergetic 60 keV x-ray source ((μx)_{mono}), corresponding to the mean energy of the XVI x-ray tube (Spezi *et al* 2009). Each pixel value of attenuation from the two sources were then linked, and a second order polynomial was fitted to the data:

$$(\mu x)_{\text{mono}} = p_0 + p_1(\mu x)_{\text{poly}} + p_2(\mu x)_{\text{poly}}^2$$
(13)

With the patient specific fitting parameters p_i determined from simulated data, the beam hardening correction was applied.

Projection scaling. The same scaling of projection images as described in the RTK(BS) method was applied to the the comprehensively corrected projection images prior to reconstruction.

Reconstruction. Reconstruction was performed with RTK employing the built-in truncation correction, using the same parameters as described previously.

2.2. Image quality evaluation

To assess the image quality of the different CBCT reconstructions, the rCT image were used as the gold standard image for each patient. Visual inspection of the reconstructed CBCT images was performed, and difference images were calculated to assess the remaining discrepancies between the corrected CBCT images and the rCTs. Furthermore, so-called calibration plots were created where the HU values in the rCT volume also covered by the CBCT reconstructions were binned in 50 equidistant bins. The median CBCT HU value in all voxels corresponding to the CT voxels in each bin was then plotted against the median CT value in the respective bin. Bins with less than 100 voxels in the CT image were discarded. To suppress effects of misalignment between the CT and CBCT images (such as positions of major vessels in the lungs, or minor positional changes of the chest wall or mediastinum), the RTK(All) reconstruction for each patient was deformably registered to the rCT image using the open source software *Elastix* (Klein *et al* 2010). This registration was subsequently applied to the other three CBCT reconstructions to ensure alignment between the four CBCT reconstructions.

As a quantitative measure of the image quality, the average HU error over the entire reconstructed CBCT volume was calculated. This is defined as

$$V_{\text{err}} = \sqrt{\text{mean}([\text{HU}_{\text{CT}}(x, y, z) - \text{HU}_{\text{CBCT}}(x, y, z)]^2)}$$
(14)

in accordance with (Bootsma *et al* 2015). Here, $HU_{CT}(x, y, z)$ denotes the HU of the rCT scan in voxel (*x*, *y*, *z*), while $HU_{CBCT}(x, y, z)$ denotes the HU in the CBCT image of the same voxel. The average HU error was calculated after 3D median filtering of the CBCT images using a $5 \times 5 \times 5$ voxel kernel to suppress the additional image noise in the corrected reconstructions.

2.3. Computer hardware

MC simulations were carried out on our computer cluster made from 24 Intel Xeon X5650 2.66 GHz CPUs. Each CPU has 6 cores and uses hyperthreading to allow a total of 288 jobs to be processed in parallel. 16 GB RAM is available on 12 compute modules, each holding 2 CPUs.

All image processing and reconstruction was performed on a laptop with an Intel Core i3-3110M 2.40 GHz CPU and 16 GB RAM. Image processing was implemented in Matlab R2014a (The MathWorks Inc, MA, USA).

3. Results

Figure 2 shows an illustrative example of axial and coronal CBCT reconstructions for patient 3. From these images, it is evident that the reconstructed CBCT HU values are too high in the lung tissues, while being too low in the tumour and bony structures on both the XVI and RTK(XVI) reconstructions. When the MC based scatter correction is applied, the HU values in the lung tissue are improved, but too low CBCT HU values in the tumour and mediastinum remain. This remaining discrepancy is resolved when all corrections are applied in the RTK(All) reconstruction. A slight overcorrection of the bony structures is observed in the RTK(All) reconstruction. Furthermore, contrast is improved and gradients between different gray values are steeper when all corrections are applied. Example images of the remaining four patients are shown in appendix B.

The calibration plots in figure 3 show the CBCT HU values of the entire reconstructed CBCT volume as a function of the reconstructed rCT HU values for all five patients. These plots emphasize that the XVI and RTK(XVI) reconstructions overestimate the CBCT HU values in the lung tissue, and underestimate the CBCT HU values for the bony structures.



Figure 2. Example images for patient 3, illustrating that all corrections are required to recover HU values in the tumour and soft tissues. The example axial and coronal images are shown with a display range of $(-250\ 1400)$, to show contrast in the lung tissue as well as soft tissue. Difference images are shown with a scale of $(-500\ 500)$. Blue colours represent too high CBCT HU values, while red colours are indicative of too low CBCT HU values compared to the rCT images.

The main difference between the XVI and RTK(XVI) reconstructions is the high CBCT HU values, where the RTK(XVI) reconstruction results in a closer match with the rCT HU values than the XVI reconstruction.

The RTK(BS) reconstructions provide a better HU match at low HU values corresponding to the lung tissue, but tends to underestimate the CBCT HU values of the soft and bony tissues. This is most pronounced for patients 1, 2, and 4, while patients 3 and 5 are nearly matched to the CT HU by the RTK(BS) reconstruction. However, when looking at the example reconstructions of patients 3 and 5 in figures 2 and B4 in the appendix, local discrepancies remain in the RTK(BS) reconstruction in the superior part of the reconstructed FOV.

When all corrections are applied in the RTK(All) reconstructions, the best one-to-one correspondence between the CBCT HU and the rCT HU values of the different reconstructions is found. For patient 4, figure 3 shows large CBCT HU variation in the high HU region (bony structures). This is caused by improper registration of a shoulder, and is not an effect of the



Figure 3. Calibration plots for the five lung patients displaying the median CBCT HU value as a function of the median CT HU value in 50 equidistant bins for each of the four reconstruction methods. Error bars show the 25 and 75% quartiles of the CBCT HU values in each bin. The large discrepancies at high HU values for patient 4 is due to improper registration of a shoulder.

artefact corrections. Due to the low number of voxels containing high HU values, this mismatch is very pronounced in the calibration plots.

To quantify the differences in CBCT HU values, the total volume error was calculated for all reconstructions of the five patients as shown in table 2. The overall trend is that the XVI reconstruction has the largest total volume error, with a slight decrease obtained by the RTK(XVI) reconstruction. The RTK(BS) reconstruction reduces the error further, but for four of the five patients, the lowest HU error is found in the RTK(All) reconstruction.

	$V_{ m err}$ (HU)			
	XVI	RTK(XVI)	RTK(BS)	RTK(All)
Pt 1	179	171	140	106
Pt 2	163	144	156	122
Pt 3	143	133	100	97
Pt 4	159	150	125	109
Pt 5	144	153	94	108
Mean(std)	157(15)	150(14)	123(26)	108(9)

Table 2. Total volume error for the five patients calculated according to (14).

3.1. Calculation time

Monte Carlo simulations were performed on the computer cluster in less than 2.5 h per patient. With the simulated images available, the projection image processing and reconstruction was performed in 10 min for the each patient for the RTK(All) reconstruction. Image processing and reconstruction was performed on the laptop PC. The extended image processing and reconstruction time could be reduced through algorithm optimisation as well as utilisation of a higher performance computer.

4. Discussion

CBCT reconstructions of five lung scans showed that CBCT image quality can be brought much closer to CT image quality through comprehensive artefact correction. In the present study, only postprocessing of raw projection data from our clinical database was performed. This emphasizes that the image quality improvements are achievable in any clinic that has stored their original projection data, without having to acquire new images or change their acquisition protocols.

This clinically oriented study found largest improvements in image quality when all corrections were applied, with the highest importance of an accurate body scatter correction method in agreement with a previous phantom study on a bench top CBCT scanner (Sisniega *et al* 2015). Calibration plots in figure 3 show the RTK(All) CBCT HU values scattered closely around the CT HU values, indicating that the remaining HU discrepancies after artefact corrections are ascribed mainly to image noise, as well as residual mismatch between the CBCT and rCT reconstructions after deformable registration. It is a weakness of the calibration plot analysis that some bins have a low number of voxels included, and thus become very prone to image noise and improper registration. The calibration plots must therefore be considered in relation to the example images, where the HU accuracy can be assessed in relation to the position in the image. The same sensitivity to image noise and improper registration is found in the total volume errors presented in table 2, which explains why what appears to be major changes in HU accuracy in the example images show up as only modest changes in the root mean square error metric.

The deformable registration itself is a potential source of bias favouring the RTK(All) correction on which the deformable registration was performed. We did however not observe major changes when initially investigating the registration accuracy of the different reconstructions, and the same registration was used for all reconstructions of the same patient to eliminate effects of small variations between individual deformable registration of the CBCT images in the comparison. Small anatomical variations between the rCT and CBCT reconstructions were still present although reduced through the deformable registration.

The main cost of the improved HU correspondence is an apparent increase in image noise. The clinical images contains statistical Poisson noise, and when the proposed image corrections are applied, signal is subtracted from the clinical images. This process leads to a decreased signal to noise ratio in the projections, resulting in decreased signal to noise in the reconstructed images. Other studies have proposed the use of iterative reconstructions to combat the statistical noise (Sidky and Pan *et al* 2008, Tang *et al* 2009), and the noise could also be reduced by a post-reconstruction filter (e.g. median filtering).

In CBCT imaging of the lungs, cardiac and respiratory motion cause blurring and streaking in reconstructed 3D images. These motion related artefacts are not considered in the present study, but at the same time they are not believed to cause major degradation of the effect of the proposed corrections. The image lag and detector scatter corrections simply correct for effects in the detector which are not affected by motion, while the patient specific correction for body scatter is a spatially slowly varying function. This means that the small changes in anatomy related to the breathing motion will not have a major effect on the body scatter correction. The motion itself was not considered in the presented correction methods, but motion artefacts can be reduced through iterative reconstruction methods (Leng *et al* 2008) or motion compensated reconstruction (Rit *et al* 2009). In addition to the motion induced artefacts, streaks arise from the undersampled CBCT acquisitions, which do not fulfil Tuy's data sufficiency condition due to the circular orbit of the gantry mounted CBCT scanner (Tuy 1983).

While the present study relies on Monte Carlo based body scatter corrections, alternative options have been proposed in the literature. Suggestions of adding an anti scatter grid to the detector (Sisniega *et al* 2013, Stankovic *et al* 2014) or a stationary or moving beam blocker close to the source (Wang *et al* 2010, Niu and Zhu 2011, Ouyang *et al* 2013, Ritschl *et al* 2015) have shown promising results, but post-processing such as the algorithmic methods proposed by (Bertram *et al* 2005, Zhao *et al* 2015) or MC based methods (Poludniowski *et al* 2009, Bootsma *et al* 2015, Xu *et al* 2015) have the advantage of being able to retrospectively correct for body scatter in images of patients who already completed treatment. This makes the algorithmic or MC based approaches such as the present model more appealing for doing retrospective research on larger patient cohorts.

The scaling factor K applied prior to reconstruction of the RTK(BS) and RTK(All) was determined from the MC simulations of primary and scattered radiation. The same scaling factor could potentially be measured on the XVI system as the open field signal, but this is not a feasible option for retrospective data analysis as the acquisition protocols may have changed. Furthermore, we have observed some variations in the output of the x-ray source when using the same preset on the same unit, and hence we favour the MC based estimation of K.

A different way of obtaining accurate HU in CBCT images has been proposed by Yang *et al* (2007) and Zhang *et al* (2015) where the planning CT is deformably registered to the daily CBCT. This approach has the strength of providing true HU values as long as the anatomy does not change too much, but the accuracy of the method depends on the accuracy of the deformable image registration which is difficult to validate. Furthermore, anatomical changes are frequent in lung cancer patients (Møller *et al* 2014), and it is not clear how the deformable image registration handles the appearance or disappearance of atelectasis or pleural effusion. While the proposed MC based scatter correction based on the planning CT image will also be affected by these anatomical changes, the scattered radiation will be less sensitive to anatomical changes than the primary radiation. In the same way, small variations in patient setup between planning CT and CBCT acquisition might have a slight detrimental effect on the scatter correction accuracy. Although the present study does not correct for

these setup variations, the smooth variation in the scatter signal as well as small setup corrections (≤ 1 cm) means that this is not expected to be a major problem.

The relative effect of each of the proposed artefact correction steps was found to be related to the individual patient rather than a common measure. The present study did not have the power to fully investigate the image quality improvement found from all potential combinations of the artefact corrections, which requires many more patients to provide a statistical measure of whether all corrections are required to obtain a significantly better image quality.

It is noted that a recent upgrade of the XVI software (R5.0) contains so-called HU-calibration which was not used in the clinical reconstructions in the present work. This clinically available HU calibration relies on post-reconstruction linearisation of the measured grey values as measured from a phantom. For patient sizes the same as the calibration phantom this method may be relatively accurate. We do however note that a change in body scatter will cause a change in HU even after calibration, and hence the method remains to be proven accurate for patients of different sizes.

How to best evaluate image quality remains an open question with many answers depending on the intended use of the images studied. Phantom studies have the strength of providing a ground truth image since the geometry and material composition of the phantom is known, but phantoms suffer from non-realistic discontinuous density interfaces and truly homogeneous materials, which are never found in patients. Furthermore, clinical CBCT imaging of lung cancer patients includes respiratory and cardiac motion far more complex than what can be simulated by 4D phantoms. Therefore, we believe that patient imaging can only truly be optimised on patient images.

The main use of CBCT image is for image guidance, and it is well established that even undersampled CBCT images can provide accurate image registration against the planning CT image (Westberg *et al* 2010). The main driver behind the quest for improved CBCT image quality is therefore not the improvement of image registration accuracy, but rather the potential extended uses of CBCT images acquired on a daily basis. In this study, a very generic image quality measure was chosen, namely the HU resemblance of CBCT images to reference CT images. We believe that if CBCT images can provide proper HU values without severe artefacts, the potential routine uses of such CBCT images include dose calculations for treatment verification, dose accumulation, and treatment adaptation, but also the extraction of anatomical biomarkers during the fractionated treatment course which might allow a much higher degree of personalised radiotherapy than what can be offered today.

5. Conclusion

A comprehensive artefact correction method for clinical CBCT images of the chest has been demonstrated to improve CBCT-CT HU correspondence for five lung cancer patients with CBCT images acquired for IGRT. This study shows the first clinical results on how sophisticated body scatter corrections do not suffice in realising the best image quality. To realise the best HU correspondence, all the proposed corrections must be applied in conjunction. No patient specific optimisation of the artefact corrections was applied, and the artefact correction methods work as retrospective processing of the clinical projection data already available. With the improved CBCT image quality, CBCT images might have the potential to be used for dose calculation and accumulation, plan adaptation, and biomarker extraction in the same way as CT images.

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Appendix A. Details of Monte Carlo simulation of the XVI source

This appendix describes the simulation setup used to simulate the XVI source in the present work. Geometrical details are estimated from information found in the technical data x-ray tube housing assembly document from Dunlee, describing the DA10 series tube housing (available at www.dunlee.com/resources/category/1/1/2/3/documents/DA%2010%20 Series%200309%20Dunlee.pdf on 18 April 2016), and the Technical data DU 694 x-ray tube document (www.dunlee.com/resources/category/1/2/5/5/images/DU%20694%201205.pdf, available on 18 April 2016), as well as the XVI R4.5 and R5.0 instructions for use by Elekta Ltd. Furthermore, non-destructive measurements of the x-ray source assembly was performed. It is noted that the described simulation geometry has only been estimated from measurements and information in the provided sources, and validated to give simulations which are similar to measured data. The geometry is thus not an accurate picture of the x-ray source assembly, but rather an empirical estimate which provides simulation results with sufficient accuracy for the intended scatter correction purpose. While some numbers are given with high accuracy in the following description, this reflects only the numbers which have been used in the actual simulation and not the accuracy with which the details are known. Most of the geometry specifications have been calculated based on specifications of field size etc.

Figure A1 shows a schematic overview of the Monte Carlo simulation setup when simulating the XVI source. The source was simulated as a compiled BEAMnrc source input to egs_cbct using a photon low energy threshold of 1 keV and an electron low energy threshold of 512 keV (total energy).

A.1. X-ray target

The x-ray target was simulated using the XTUBE component module in BEAMnrc, with the target defined as a 5 mm thick alloy composed of 90% tungsten and 10% rhenium by weight, giving a mass density of 19.4 g cm⁻³. The target alloy was mounted on a molybdenum holder. The target angle was 17.5°, and the *z*-extend of the target was estimated to 1 cm. The centreline of the x-ray target was defined as the reference plane with z = 0.

A.2. Tube exit window

The tube window was simulated using the PYRAMIDS component module to define the aperture of the Dunlee x-ray tube housing. The window was positioned at an estimated z = 5.73 cm, with a square aluminium window of 1.6 mm thickness and side length 2.64 cm. The Al window was placed in the opening of a 3.2 mm lead block, and the opening was focused towards the focal point of electrons on the x-ray target.



Figure A1. Schematic overview of the Monte Carlo simulation setup for the XVI tube. Figure not to scale.

A.3. Filtration

Additional filtration was simulated using the SLABS component module placed immediately after the tube exit window at z = 6.05 cm. The filtration was composed of 0.1 mm copper and 2 mm aluminium.

A.4. Primary collimator

The primary collimator in the XVI tube assembly was simulated using the PYRAMIDS component module. An unfocused square aperture with an estimated side length of 3 cm was defined in a 3.2 mm thick lead block placed at z = 6.5 cm.

A.5. S20 collimator

The S20 collimator cassette used in the present study was simulated using the SLABS, PYRAMIDS, and SLABS component modules. The S20 collimator is constructed as a lead aperture placed between two sheets of a transparent plastic material. This plastic was simulated as PETG ($C_{10}H_8O_4$, density 1.27 g cm⁻³). The first PETG slab was placed at z = 19.89 cm, and with an estimated thickness of 1 mm. This was followed by the square collimator specified using the PYRAMIDS component module, creating a 5.67 cm square opening in a 3.2 mm thick lead block. The aperture was then followed by another PETG sheet of 1 mm thickness.

A.6. FO filter

The F0 filter simply consists of two plastic sheets with air in between. This was simulated using the SLABS component module with three layers placed at z = 21 cm. The first layer was 1 mm PETG followed by 3 cm air, and finally a 1 mm PETG layer.



Appendix B. Example images of patients 1, 2, 4 and 5

Figure B1. Example images for patient 1. The irregular shape of the axial CBCT FOV is a result of the deformable image registration. The example axial and coronal images are shown with a display range of $(-250\ 1400)$, to show contrast in the lung tissue as well as soft tissue. Difference images are shown with a scale of $(-500\ 500)$.



Figure B2. Example images for patient 2. The example axial and coronal images are shown with a display range of $(-250\ 1400)$, to show contrast in the lung tissue as well as soft tissue. Difference images are shown with a scale of $(-500\ 500)$.



Figure B3. Example images for patient 4. The example axial and coronal images are shown with a display range of $(-250\ 1400)$, to show contrast in the lung tissue as well as soft tissue. Difference images are shown with a scale of $(-500\ 500)$.



Figure B4. Example images for patient 5. The example axial and coronal images are shown with a display range of $(-250 \ 1400)$, to show contrast in the lung tissue as well as soft tissue. Difference images are shown with a scale of $(-500 \ 500)$.

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